been used. This is adequate to keep molds from appearing and seems not to give bad effects on the flies.

Bridges, C. B. On the seeding of culture with yeast.

On some five occasions the method of seeding the surfaces of culture bottles by spraying with very thin yeast suspension (see DIS-6;66) has been hopefully out into practice and then aband-

oned in favor of seeding with one, or better two fat drops of thick yeast suspension. Always the failures of pair cultures to produce offspring became so large as to seriously hinder the experimental breeding. The main advantage of the spraying was the suppression of mold growth - and this is now better accomplished by moldex.

Bridges, C. B. On distinguishing larvae for salivary preparations.

The notes by Beadle (Am. Nat.71: 277; DIS-6:24), Hoover (DIS-6:24) and Brehme DIS-8: -) show how the distinction between the yellow of normal malphigian tubules and the colorless or paler tubules

associated with certain light eye-colors (notably w and lt but also cm, g² p⁰ and ca) and the brown color of the mouth parts of yellow larvae, can be used to select larvae of the type desired for salivary preparations. A survey of our balancers shows that a few of them are especially useful for general use in balancing any mutant whose salivaries might need investigation. For chromosome I, these are: Cl, y Hw (carrying y and g²) dl-49, m² g⁴ (carrying g⁴, already recommended by Hoover) and dl-49, w lz⁸. For chromosome II, the best is Cy, al² lt³ L⁴ sp² (carrying lt³ of Beadle) and for chromosome III, the best is Payne, Dfd ca. These balancers are the best of the ClB, dl-49, Cy and Payne varieties, and should be kept on hand and favored in stock making.

For second and third chromosome aberrations, use can be made of the dominant eye-color pg which has pale tubules (perhaps pg/pg are better). First cross to T(Y;2;3)pg, pick out the T(2;3)pg aberration of and cross to pp ? 2, using the normal yellow tubuled larvae.

A second and third chromosome method needing no distinguishing of larvae, except the easy one of femaleness, has been the use (by Schultz and myself) of T(Y;2;3)I. Cross the female bearing the aberration to \$ males carrying T(Y;2;3)I, pick out sons carrying the aberration (all are T(Y;2;3)I and cross to any standard female. All daughters are heterozygotes for the aberration.

Buzzati-Traverso, A.

Method for making salivary gland chromosomes
permanent smearing.

I found very convenient for making permanent salivary gland chromosome smears the following method: (1) Dissect as usual the larva and leave the salivary gland in normal aceto carmine till well stained; (2) pass the gland

to the slide and take off all the aceto carmine which might

be on the slide, avoiding the gland to dry; (3) let two or three drops of the following liquid fall on the gland; l part Faure's liquid (50cc. of distilled water / 50cc. of chloral hydrate / 20cc. of glycerine / 30cc. of arabic gum) mixed with 2 parts of normal aceto carmine (the proportion 1:2 should be changed slightly to fit the different materials); (4) put the cover slip on and smear as usual, leaving the liquid in excess to dry around the edges of the cover slip. The slide will be dry enough to be used in a few hours, and keeps in very good conditions practically indefinetely.

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Just, G. and F. Steiniger. Food.

The marketable fruitmarmalades, which could be used as a convenient Drosophila food, in Germany nowadays contain reme-

dies for preventing fermentation, and the food becomes very acid in short time. These marmalades are to be used only with great cautiousness.

Luers, H. The use of the dominant bobbed-muta-nant Bobbed in the Y-chromo-some of D. funebris in gene-tic experiments

some of D. funebris in genetic experiments.

rays in the Y-chromosome
of Drosophila funebris
(Bb). Some properties of this new mutation make it useful in some genetic

experiments. (1) BbX has a markedly slower development. Since it is present in males only, it facilitates the obtaining of virgin females from mass-cultures. (2) Bby is a good marker of the Y-chromosome. (3) Bby enhances non-disjunction of the X-and Y-chromosomes, and facilitates the obtaining of XO- and XYY- o and of XXX-, XXY- and XXYY- 22.

Medvedev, N.N. How to make Drosophila larvae immobile for a short time.

In order to carry out transplantation experiments on Drosophila lar-vae it is necessary to make them for some time immobile. Beadle and

Ephrussi in their work use the current method of etherization. This treatment, however, is very undesirable, because it makes transplantation experiments tedious by themselves still more difficult, especially in the case when they are carried out by but one person.

For this purpose the author successfully uses a very simple method. After placing a larvae on the slide where we are going to perform the transplantation, it is quite sufficient to press it gently with a piece of filter paper and to roll it over a few times around its longitudinal, axis. After this simple manipulation the larva becomes immobile for a time sufficiently long to perform a transplantation.